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(54) Title: DETECTION METHOD FOR C-RAF-1	GENE	

(57) Abstract

The present invention relates to: (1) a method of identifying an individual at an increased risk for developing cancer, (2) a method for determining a prognosis in patients afflicted with cancer, and (3) a method for determining the proper course of treatment for a patient afflicted with cancer; comprising: amplifying a region of the c-raf-1 gene.

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DETECTION METHOD FOR C-RAF-1 GENES

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Field of the Invention

The present invention relates to (1) a method of identifying an individual at an increased risk for developing cancer, (2) a method for determining a prognosis of patients afflicted with cancer, and (3) a method for determining the proper course of treatment for a patient afflicted with cancer.

Background Information

Lung cancer claims more lives in the United 20 States than any other neoplasm (R.S. Finley, Am. Pharm. NS29, 39 (1989)), and of the various forms lung adenocarcinomas have one of the worst prognoses (T.P. Miller, Semin. Oncol. 17, 11 (1990)). The incidence of adenocarcinoma of the lung (ACL) in the United States is 25 also quickly rising (I. Linnoila, Hematol. Oncol. North. Am. 4, 1027 (1990); J.B. Sorensen, H.H. Hansen, Cancer Surviv. 8, 671 (1989)). In order to gain insight into this complex and deadly disease, a model system for its study has been developed. For such a model to provide 30 clinically relevant data several criteria must be met. The tumors produced should be histologically equivalent to their human counterparts, tumor induction must be reliable and reproducible, and the numbers generated must be great enough to provide statistical significance. To satisfy 35 these conditions a system has been created which uses two inbred mouse strains (NFS/n and AKR) along with

transplacental exposure to the potent carcinogen N-ethyl-N-nitrosourea (ENU) and promotion with the antioxidant butylated hydroxytoluene (BHT). The resulting tumors were examined for altered expression or structural mutations of genes implicated in lung tumor development such as ras, myc, and raf oncogenes (C.D. Little et al., Nature 306, 194 (1983); P.E. Kiefer et al., Cancer Res., 47, 6236 (1987); E. Santos et al., Science 223, 661 (1984); S. Rodenhuis, N. Engl. J. Med. 317, 929 (1987); M. Barbacid, Eur. J. Clin. Invest. 20, 225 (1990); U.R. Rapp et al., J. Int. Assoc. for the Study of Lung Cancer 4, 162 (1988); M.J. Birrer et al., Ann. Rev. Med. 40, 305 (1989); G. Sithanandam et al., Oncogene 4, 451 (1989)).

raf proto-oncogenes are evolutionarily highly 15 conserved genes encoding cytoplasmic serine/threonine specific kinases, which function in mitogen signal transduction (reviewed in U.R. Rapp et al., The Oncogene Handbook, T. Curran et al., Eds. (Elsevier Science 20 Publishers, The Netherlands, 1988), pp. 115-154; U.R. Rapp, Oncogene 6, 495 (1991)). The three known active members in the <u>raf</u> family encode phosphoproteins of similar size (72/74 kD for Raf-1; 68 kD for A-Raf-1, and 74 kD for B-Raf (U.R. Rapp et al., in Retroviruses and 25 <u>Human Pathology</u>, R. Gallo et al., Eds. (Humana Press, Clifton, New Jersey 1985), pp. 449-472; T.W. Beck et al., Nucleic Acids Res. 15, 595 (1987); G. Sithanandam et al., Oncogene 5, 1775 (1990))). Raf-1 was first identified as the cellular homologue of v-raf (H.W. Jansen et al., 30 <u>Nature</u> 307, 218 (1984)), the transforming gene of 3611 MSV (U.R. Rapp et al., <u>J. Virol.</u> 45, 914 (1983); U.R. Rapp et al., Proc. Natl. Acad. Sci. USA 80, 4218 (1983)). Amino acid comparisons of raf family genes shows three conserved regions [CR1, CR2, CR3] (T.W. Beck et al., Nucleic Acids 35 <u>Res.</u> 15, 595 (1987)); CR1 is a regulatory region surrounding a Cys finger consensus sequence, CR2 is a serine/threonine rich region, and CR3 represents the

kinase domain. Raf-1 has been mapped to chromosome 3p25 in humans (S.J. O'Brien et al., Science 223, 71 (1984)), and this region has been found to be frequently altered in small cell lung carcinoma (SCLC) (J. Whang-Peng et al., Cancer Genet. Cytogenet. 6, 119 (1982); J.M. Ibson et al., J. Cell. Biochem. 33, 267 (1987)), familial renal cell carcinoma (A.J. Cohen et al., N. Engl. J. Med. 301, 592 (1979); G. Kovacs et al., Int. J. Cancer 40, 171 (1987)), mixed parotid gland tumors (J. Mark et al., Hereditas 96, 141 (1982)), and ovarian cancer (K. Tanaka et al., Cancer Genet. Cytogenet. 43, 1 (1989)).

Raf genes are differentially expressed in various tissues (S.M. Storm et al., Oncogene 5, 345 15 (1990)). c-raf-1 has been found to be expressed ubiquitously, though absolute levels vary between tissues. A-raf-1 is present predominantly in the urogenital tissues, whereas B-Raf is most abundant in cerebrum and testis. The ubiquitous c-Raf-1 kinase is regulated by 20 tyrosine and serine phosphorylations that result from activated growth factor receptor kinases (D.K. Morrison et al., Cell 58, 648 (1989); D.K. Morrison et al., Proc. Natl. Acad. Sci. USA 85, 8855 (1989); K.S. Kovacina et al., J. Biol. Chem. 265, 12115 (1990); P.J. Blackshear et 25 al., <u>J. Biol. Chem.</u> 265, 12131 (1990); M.P. Carroll et al., <u>J. Biol. Chem.</u> 265, 19812 (1990); J.N. Siegel et al., J. Biol. Chem. 265, 18472 (1990); B.C. Turner et al., Proc. Natl. Acad. Sci. USA 88, 1227 (1991); M. Baccarini et al., EMBO J. 9, 3649 (1990); H. App et al., Mol. Cell. 30 Biol. 11, 913 (1991)). Raf-1 operates downstream of Ras in mitogen signal transduction as judged by experiments using antibody microinjection (M.R. Smith et al., Nature 320, 540 (1986)), c-raf-1 antisense expression constructs (W. Kolch et al., Nature 349, 426 (1991)), dominant 35 negative mutants (W. Kolch et al., Nature 349, 426 (1991)), and Raf revertant cells. Studies with NIH3T3 cells and brain tissue demonstrated that mitogen treatment

induces Raf-1 kinase activity and causes a transitory relocation of the active enzyme from the cytoplasm to the nucleus and perinuclear area (Z. Olàh et al., Exp. Brain. Res. (in press); U.R. Rapp et al., in Cold Spring Harbor Symposia on Quantitative Biology, Vol. LIII, Eds. (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY 1988) pp. 173-184).

Raf-1 coupling has been examined in more than a dozen receptor systems and all strong mitogens stimulated Raf-1 kinase activity (U.R. Rapp, Oncogene 6, 495 (1991); D.K. Morrison et al., Cell 58, 648 (1989); D.K. Morrison et al., Proc. Natl. Acad. Sci. USA 85, 8855 (1989); K.S. Kovacina et al., J. Biol. Chem. 265, 12115 (1990); P.J. Blackshear et al., J. Biol. Chem. 265, 12131 (1990); M.P. Carroll et al., J. Biol. Chem. 265, 19812 (1990); J.N. Siegel et al., J. Biol. Chem. 265, 18472 (1990); B.C. Turner et al., Proc. Natl. Acad. Sci. USA 88, 1227 (1991); M. Baccarini et al., EMBO J. 9, 3649 (1990); H. App et al., Mol. Cell. Biol. 11, 913 (1991)), and this stimulation correlated with an increase in Raf-1 phosphorylation leading to a shift in apparent molecular weight.

SUMMARY OF THE INVENTION

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It is an object of this invention to provide a method of identifying an individual at an increased risk for developing cancer.

It is another object of this invention to provide a method for determining a prognosis in patients afflicted with cancer.

35 It is a further object of this invention to provide a method for determining the proper course of treatment for a patient afflicted with cancer.

WO 93/06248 PCT/US92/07817

Further objects and advantages of the present invention will be clear from the description that follows.

In one embodiment, the present invention relates to a method of identifying an individual at an increased risk for developing cancer, comprising:

amplifying a region of the c-raf-1 gene; analyzing products of the amplification for evidence of mutation; and

10 classifying an individual having one or more mutations in the region as having an increased risk for developing cancer.

In another embodiment, the present invention 15 relates to a method for determining a prognosis in patients afflicted with cancer, comprising:

amplifying a region of the c-raf-1 gene; analyzing products of the amplification for evidence of mutation; and

classifying patients having no mutation in said region as being less likely to suffer disease relapse or having an increased chance of survival than those patients having one or more mutations in said region.

In a further embodiment, the present invention relates to a method for determining the proper course of treatment for a patient afflicted with cancer, comprising:

amplifying a region of the c-raf-1 gene;

analyzing products of said amplification for evidence 30 of mutation;

identifying a patient having at least one mutation in said region, which patient may require treatment proper for patients having a lesser chance of survival or decreased time to relapse; and

identifying a patient lacking mutations in said region, which patients may require treatment proper for

PCT/US92/07817 WO 93/06248

-6-

patients having a greater chance of survival or being less likely to suffer disease relapse.

BRIEF DESCRIPTION OF THE DRAWINGS

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Figure 1. Effect of BHT promotion on ENU tumorigenesis in NFS/n x AKR mice. The X-axis represents percent tumor induced mortality within each group, and the Y-axis reflects age in weeks. All animals were exposed to 10 ENU transplacentally at a dose of 0.5mM/Kg of mother's body weight on day 16 of gestation (presence of vaginal plug was scored as day one). At two weeks of age mice were weaned into two separate groups and separated by sex. Both groups were housed in identical cages and supplied 15 with food (Purina Lab Chow) and water ad libitum. Beginning at three weeks of age, group 2A (0) was given weekly intraperitoneal (i.p.) injection of corn oil (0.1 ml), and group 2B (>) received weekly i.p. injections of BHT (20 mg/Kg of body weight) dissolved in corn oil. 20 Administration of BHT reduces the mean age of mortality from approximately 20 weeks to 13, and decreases the initial age of mortality. These curves are significantly different (p≤ 0.001) as judged by a 2-tailed Cox test. In both groups the rate of tumorigenesis was identical for 25 males and females.

Figure 2. Northern blot analysis of protooncogene expression levels in ENU induced tumors.

Figure 3. Diagnostic digestion of PCR amplified 30 Ki-ras genes from ENU induced tumors. Genomic DNA was isolated from a cesium chloride gradient during RNA preparations. In each case 10 ng was amplified via PCR (95°C, 5 min. followed by 35 cycles of 95°C, 1 min. → 35 55°C, 1 min. \rightarrow 72°, 1 min.) with 2 units of Taq I polymerase. The primers used (K1; 5'-AACTTGTGGTGGTTGGACCT-3'→ (SEQ ID NO:6) and K2; <= 3'-

GTCTTAGTGAAACACCTACT-5' (SEQ ID NO:7)) generate a 79 b.p. product. The primer K1 ends at codon 12 and contains a mismatch from normal mouse (Ki-ras sequence its 18th nucleotide (G \rightarrow C) creating a BstNI site (CCTGG) in 5 conjunction with a normal codon 12 (GGT). Digestion of amplified product from a normal allele with BstNI produces two products of 19 and 60 b.p., whereas a mutation in one of the first two positions of codon 12 will eliminate the BstNI site. The presence of two normal alleles results in 10 all of the product being cleaved and the presence of one mutant and one normal allele will result in only half of the product cut. In the three panels each sample was run in duplicate, uncut and cut with BstNI. F1 is DNA from an untreated NFS/n X AKR F1 mouse, and MCA5 is a murine cell line known to harbor a mutant K-ras codon 12 allele. One lymphoma (24Ly) and one cell line (117; derived from a lung adenocarcinoma) display a mutated Ki-ras codon 12 allele; however, 24Ly was a passaged tumor and examination of the original tumor showed two normal alleles indicating 20 that this mutation was acquired during passaging.

Figure 4. c-raf-1 RNAse protection analysis of ENU induced tumors. The probe used was a 32P labeled antisense transcript from the 3' non-coding region of a 25 mouse c-raf-1 cDNA to the 3' most StuI site. Hybridization of this probe with normal RNA results in a protected fragment of 1.2kb covering the region encoding the Raf-1 kinase domain. One µg of poly(A)+ RNA from each tumor and 5 μg of F1 RNA (in order to get comparable 30 signals) was hybridized for 12 hours at 52°C with 200,000 cpm of 32P labeled mouse c-raf antisense transcript. Hybrids were then digested for 30 minutes with 25 µg RNAse A and 33 units of RNAse T1 at room temperature. Digested hybrids were then incubated with 50 µg of proteinase K, 35 phenol/chloroform extracted, ethanol precipitated, and resuspended in loading dye containing 80% formamide. Samples were then run on 6% polyacrylamide denaturing

WO 93/06248 PCT/US92/07817

-8-

sequencing gels at 65 watts. Gels were vacuum dried at 80 degrees C and exposed to X-ray film. Probe is undigested probe alone; tRNA is probe hybridized to non-specific RNA; v-raf is probe hybridized to RNA from a v-raf transformed cell line and the bands detected represent single base mismatches between murine c-raf and v-raf; NFS/AKR F1 is probe hybridized with RNA from a normal (untreated) mouse; 24 LY is probe hybridized with RNA from a lymphoma; and the remaining lanes are probe hybridized with RNA isolated from lung tumors.

Figure 5. Schematic of Raf-1 protein showing sites of ENU induced mutations. CR1, CR2, and CR3 represent conserved regions 1, 2 and 3. cDNAs were made 15 from tumor derived poly(A)+ RNA using MoMuLV reverse transcriptase. Primers (MR1 sequence and MR2 sequence) encompassing a 435 base pair region c-raf were then used to amplify this region via PCR. The amplification mixture was then run on 1.7% agarose gels and the 435 bp product isolated. This isolated fragment was then treated with T4 polymerase and cloned into the HincII site of M13mp18 for 20 sequencing. Another set of primers (EMR1 sequence and EMR2 sequence) was designed containing EcoRI sites at the termini and used to amplify a 609 base pair region (encompassing the original 435 base pair region). Isolated products from these reactions were then digested with EcoRI and cloned into the EcoRI site of KS. Sequencing reactions were carried out using the Sequenase kit (USB) according to the recommended protocols for 30 single and double stranded sequencing. Sequencing reactions were run on 6% polyacrylamide denaturing gels at 65 watts. Gels were vacuum dried at 80 degrees C and exposed to X-ray film. In each case a normal allele was also sequenced along with the mutant allele.

WO 93/06248 PCT/US92/07817

-9-

Figure 6. Schematic for Identifying c-raf-1 mutations. Primers 1 and 2 are shown in SEQ ID NO:8 and SEQ ID NO:9, respectively.

DETAILED DESCRIPTION OF THE INVENTION

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The present invention relates to methods that involve amplifying a region of the c-raf-1 gene (the sequence of a mouse c-raf-1 gene is shown in SEQ ID NO:10; the nucleotide and corresponding amino acid sequence of a human c-raf-1 gene is shown in SEQ ID NO:11 and SEQ ID NO:12, respectively).

In one embodiment, the present invention relates 15 to a method of identifying an individual at an increased risk for developing cancer (preferably, lung cancer, Tcell lymphomas, renal cell carcinoma, ovarian carcinoma, and mixed parotid gland tumors) comprising: amplifying a region (preferably by using the polymerase chain reaction 20 method(PCR) or by cloning techniques) of the c-raf-1 gene of the individual (SEQ ID NO:11)(in one preferred embodiment, the region encodes amino acids 514 to 535 of SEQ ID NO:12); analyzing products of the amplification for evidence of mutation (preferably by DNA sequencing of the 25 region) and classifying an individual having one or more mutations in the region as having an increased risk for developing cancer. In one prefered embodiment, the region encodes amino acids 500 to 550 of SEQ ID NO:12 or amino acids 450 to 630 of SEQ ID NO:12. In another prefered 30 embodiment, the PCR method employs a primer comprising the sequence shown in SEQ ID NO:7 and a primer comprising the sequence shown in SEQ ID NO:8. In another prefered embodiment, the method comprises the steps shown in Figure 6.

In another embodiment, the present invention relates to a method for determining a prognosis in a patient afflicted with cancer (preferably, those cancers

PCT/US92/07817 WO 93/06248

listed above). The method comprises: amplifying the region of the c-raf-1 gene as described above; analyzing products of the amplification for evidence of mutation as described above; and classifying a patient having no 5 mutation in the region as being less likely to suffer disease relapse or having an increased chance of survival than a patient having one or more mutations in the region.

In another embodiment, the present invention 10 relates to a method for determining the proper course of treatment for a patient afflicted with cancer (preferably, those cancers listed above), comprising: amplifying a region (described above) of the c-raf-1 gene as described above; analyzing products of the amplification for 15 evidence of mutation as described above; identifying a patient having at least one mutation in the region, which patient may require treatment proper for patients having a lesser chance of survival or decreased time to relapse; and identifying a patient lacking mutations in the region, 20 which patients may require treatment proper for patients having a greater chance of survival or being less likely to suffer disease relapse.

Administration of therapeutic agents (cytotoxic 25 or cytostatic) tailored to recognize the mutant Raf-1 protein but not normal Raf-1 could specifically target tumor cells for death of growth inhibition. Such agents could be comprised of cytotoxic T-cells, antibodies, and/or specifically designed chemical compounds.

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The following Examples demonstrate consistent point mutations of the c-raf-1 proto-oncogene, within a small region of the kinase domain, in a mouse model for chemical tymor induction. This is the first demonstration 35 of point mutated <u>raf</u> genes <u>in vivo</u>, and the first isolation of activating in vivo point mutations in the kinase domain of a proto-oncogene. The tumors examined

WO 93/06248 PCT/US92/07817

-11-

show a selective specificity for Raf-1 mutations as another family of genes, the <u>ras</u> proto-oncogenes which are frequently activated by point mutation in both animal and human tumors (S. Rodenhuis et al., <u>Am. Rev. Respir. Dis.</u> 142, S27-30; T.R. Devereux et al., <u>Carcinogenesis</u> 12, 299 (1991)), is not involved.

The present invention is described in further detail in the following non-limiting examples.

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EXAMPLES

The following protocols and experimental details are referenced in the examples that follow:

15 RNA Isolation. Tumors were excised, a small portion minced in PBS (phosphate buffered saline solution) for passaging in nude mice, frozen immediately in a dry ice/ethanol bath, and stored at -70° until RNA extraction. Frozen tissues were minced on wet ice in a guanidine 20 thiocyanate buffer (4M guanidine thiocyanate 10mM EDTA, 2% N-lauryl sarcosine, 2% beta-mercaptoethanol, 10mM Tris (pH=7.6)), disrupted in a Dounce homogenizer, and extracted three times with phenol: chloroform: isoamyl alcohol (24:24:2). Supernatants were then transferred to 25 SW41 tubes, 100 μg of cesium chloride per ml was added to the supernatant which was then underlayed with one half saturated cesium chloride in 10mM EDTA (pH=7.0; index of refraction 1.3995-1.4000), and centrifuged at 25,000 rpm for 20 hours in a Sorvall SW-41TI rotor using a Beckman 30 model L5-50 ultracentrifuge. Supernatants were removed and RNA pellets dissolved in 4 ml resuspension buffer (10 mM Tris-HCl pH=7.6, 5% beta-mercaptoethanol, 0.5% N-lauryl sarcosine, 10 mM EDTA), extracted once with phenol:chloroform:isoamyl alcohol, sodium acetate added to 0.12M and RNA precipatated with two volumes ethanol at -20°C overnight. Precipitates were centrifuged at 9,000 rpm in a Sorvall SS-34 rotor for 30 minutes, and pellets

WO 93/06248 PCT/US92/07817

-12-

redissolved in RNA sample buffer (10 mM Tris pH=7.4, 1mM EDTA, 0.05% sodium dodecyl sulfate) and concentrations determined by absorbance at 260 nm. Poly (A)* RNA was isolated by binding to oligo dT cellulose columns in high salt (10mM Tris pH=7.4, 1mM EDTA, 0.05% SDS, 500mM NaCl), and eluting with RNA sample buffer heated to 40°C.

Northern Blotting. 5 μg poly(A)* RNA per lane was ethanol precipitated, desiccated, resuspended in loading buffer (20mM MOPS pH=6.8, 5mM sodium acetate, 1mM EDTA, 50% formamide, 6% formaldehyde), heated at 65°C for 5 min., quick chilled on wet ice for 10 min., and electrophoresed through a 0.7% agarose gel containing 2.2 M formaldehyde, 20mM MOPS [pH=6.8], 5mM sodium acetate, and 1mM EDTA.

15 Gels were then blotted overnight onto nitrocellulose filters via capillary transfer in 20x SSC, filters were washed in 3x SSC for 10 min. and baked at 80°C for 2 hours.

Hybridizations. Filters were prehybridized at 42°C in 5X SSC, 50% formamide, 20mM sodium phosphate pH=6.8, 200 μg/ml PVP-40, 200 μg/ml ficoll 400, 200 μg/ml bovine serum albumin, and 200 μg/ml sonicated sheared salmon sperm DNA. Blots were then hybridized with 500,000 cpm/ml of random primed ²P labeled probes overnight at 42°C in prehybridization solution with 5% dextran sulfate. Blots were washed with agitation in 2X SSC, 0.1% SDS at room temperature six times for 20 minutes each wash, then washed once at 45°C in 0.1X SSC for 15 minutes. Filters were exposed to X-AR 5 film at -70°C.

EXAMPLE 1 Tumor Induction

NFS female mice were mated with AKR males and pregnant females given a transplacental injection of 1-ethyl-1-nitrosourea (ENU) at a dosage of 0.5 mM/Kg

mother's body weight on day 16 of gestation, counting plug date as day one. ENU was chosen for tumor induction since it is a very potent direct acting carcinogen capable of modifying any base in vivo (Singer, B. et al., 1983. 5 Molecular Biology of Mutagens and Carcinogens, Plenum Press, New York). ENU alkylates all tissues with roughly the same efficiency (E. Scherer et al., Cancer Lett. 46, 21 (1989)) and has a very short half life in vivo (E.M. Faustman et al., Teratology 40, 199 (1989)) allowing 10 specific mutagenesis of tissues which are mitotically active at a particular time. NFS and AKR were chosen as parental strains based on earlier studies which showed them to be particularly susceptible to ling tumors following ENU exposure (B.A. Diwan et al., Cancer Res. 34, 15 764 (1974); S.L. Kauffman, JNCI 57, 821 (1976)). With this procedure nearly 100% of the offspring develop lung adenocarcinomas and approximately 70% develop, in addition, T-cell lymphomas with a mean latency of approximately 20 weeks. In order to achieve more rapid 20 tumor development, weanling mice were treated with weekly injections of a tumor promoter, the antioxidant butylated hydroxytoluene or BHT (20mg/kg body weight dissolved in corn oil). BHT was used as it has been demonstrated to cause lung lesions and hyperplasia when injected into mice 25 (A.A. Marino et al., <u>Proc. Soc. Exp. Biol. Med.</u> 140, 122 (1972); H. Witschi et al., Proc. Soc. Exp. Biol. Med., 147, 690 (1974); N. Ito et al., CRC Crit. Rev. Toxicol. 15, 109 (1984)). In the present system it nearly doubles the rate at which tumors develop. Figure 1 compares tumor induced mortality with age of animals for those receiving ENU alone, and those receiving ENU and promoted with BHT. These curves demonstrate that when BHT is given the mean age of tumor induced mortality decreases from approximately 20 weeks to around 12, and there is also a 35 decrease in initial latency. These curves are significantly different with a confidence limit greater than 99.99% using a 2-tailed Cox test. In addition, BHT

PCT/US92/07817 WO 93/06248

-14-

promotion, while increasing the rate at which tumors develop, does not affect the tumor spectrum.

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EXAMPLE 2

Oncogene Expression

Northern blot analysis revealed elevated levels of c-raf-1, as compared to normal tissue, in every tumor examined (Figure 2), and Western blot analysis showed that 10 protein levels correlated with message levels (U.R. Rapp et al., in Oncogenes and Cancer, S.A. Aaronson et al., Eds. (Tokyo/VNU Scientific Press, Tokyo, 1987) pp. 55-74). In addition, in cell lines derived from primary tumors, Raf-1 protein kinase activity was shown by immune-complex 15 kinase assays to be constitutive. Further analysis of other oncogenes revealed no consistent pattern of expression except for ras and myc family genes. In the case of the myc family, one member (either c-, N-, or Lmyc) was overexpressed but never more than one. For the 20 <u>ras</u> genes, at least one member (Ki-, Ha-, or N-<u>ras</u>), and often more than one, was expressed at high levels when compared with the normal tissue. In addition all oncogenes examined via Northern analysis exhibited full length, normal sized transcripts.

ras genes were considered likely candidates for mutational activation since oncogenic forms of Ki-ras have previously been observed in lung tumors (S. Rodenhuis et al., Am. Rev. Respir. Dis. 142, S27-30; T.R. Devereux et 30 al., Carcinogenesis 12, 299 (1991)) and ENU is a point mutagen (Singer, B. et al., 1983. Molecular Biology of Mutagens and Carcinogens, Plenum Press, New York). A systematic analysis of various ras codons known to be involved in oncogenic activation was therefore performed . 35 Ha-, Ki-, and N- ras were examined at codons 12, 13, and 61 for potential mutations via RNAse protection assays (R.M. Myers et al., <u>Science</u> 230, 1242 (1985); E. Winter et

al., Proc. Natl. Acad. Sci. USA 82, 7575 (1985)), PCR amplification followed by subsequent sequencing (F. Sanger et al., <u>J. Mol. Biol.</u> 13, 373 (1975)), and PCR amplification followed by diagnostic restriction digests 5 (W. Jiang et al., Oncogene 4, 923 (1989)). PCR amplification creating diagnostic enzyme sites is a very efficient way of examining alleles for mutations at known sites and involves designing a PCR primer whose 3' end lies next to and produces a novel restriction site 10 encompassing the codon of interest. Following amplification, PCR products from normal alleles will contain the new restriction site, while mutant alleles will not. Digestion of the product from tissue with two normal alleles results in all product being cut; however, 15 if one allele contains a mutation, only half of the product will be digested. Figure 3 shows the results of amplification and diagnostic digestion applied to Ki-ras codon 12 in several tumors and cell lines. The first panel is from a set of lymphomas. F1 is DNA from a normal 20 untreated mouse and both alleles are cut by BstNl, indicating the presence of two normal alleles. MCA5 is a murine cell line known to contain a Ki-ras codon 12 mutation (L.F. Parada et al., Mol. Cell. Biol. 3, 2298 (1983)), and only the amplified normal allele is cleaved. 25 Of the five tumors shown in the second panel, one shows a mutant Ki-ras allele. The next panel shows some of the lung tumors tested and none of them exhibit a mutant allele, and the final panel shows tumor derived cell lines. The first three are from lymphomas and the last 30 three from lung adenocarcinomas. One lung tumor line (#117) has a Ki-ras 12 mutation that was not present in the primary tumor but came up upon transplantation. This analysis has been performed with Ki, Ha and N-ras genes at codons 12 and 61. Of all the tumors and cell lines 35 examined by this method for mutations of the three ras genes at codons 12 and 61, the two shown here were the only ones detected. Examination of codon 13 was done by

PCR amplification of genomic DNA surrounding codon 13
followed by cloning into KS+ (Stratagene) and double
stranded sequencing. Table I summarizes the ras mutation
data. The most notable point from this table is the

5 conspicuous lack of ras mutations in these tumors. In
fact the number of ras mutations is much lower than would
be expected for a sampling of spontaneous tumors (S.
Rodenhuis et al., Am. Rev. Respir. Dis. 142, S27-30; T.R.
Devereux et al., Carcinogenesis 12, 299 (1991); J.L. Bos,

10 Cancer Res. 49, 4682 (1989)). Having eliminated ras genes
as playing a primary role in the genesis of these ENU
induced tumors, c-raf-1 was investigated for possible
small or point mutations.

15 TABLE I
Tumors and Cell Lines Positive for ras Mutations

THIMOTS	and Cell I	<u> Lines E</u>	ositive	TOL LAS	Flucacio.	<u></u>
TUMOLU					Codon	61
		Cell	Tumors	Cell Lines	Tumors	Cell Lines
	Tumors	Titles	Tuniore		- /	016
Ha-ras	0/10	0/6	0/6	0/2	0/10	0/6
	, •	1/6	0/6	0/2	0/6	0/2
K1/Tas			0/6	0/2	0/10	0/6
N-ras	0/10	0/6	0/6	0/2	• • • • • • • • • • • • • • • • • • • •	
	Ha-ras Ki/ras	Tumors Ha-ras 0/10 Ki/ras 1*/10	Codon 12 Cell Tumors Lines Ha-ras 0/10 0/6 Ki/ras 1*/10 1/6	Codon 12 Codon Cell Tumors Lines Tumors Ha-ras 0/10 0/6 0/6 Ki/ras 1*/10 1/6 0/6	Codon 12 Codon 13 Cell Cell Tumors Lines Tumors Lines Ha-ras 0/10 0/6 0/6 0/2 Ki/ras 1*/10 1/6 0/6 0/2	Cell Tumors Lines Tumors Lines Tumors Ha-ras 0/10 0/6 0/6 0/2 0/10 Ki/ras 1*/10 1/6 0/6 0/2 0/6

This was a second passage tumor in which the original tumor did not contain a Ki-ras mutation.

Table I: Summary of mutation analysis for Ha-, Ki-, and N-ras at codons 12, 13, and 61. Each box displays the number of mutations detected, over the number of tumors and tumor derived cell lines examined via RNAse protection, sequencing or diagnostic digestion, for each of the nine codons.

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EXAMPLE 3 Mutations in Raf-1

Since no point mutations had been described for <u>raf</u> genes in vivo, as had been for the ras genes (E.

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Santos et al., <u>Science</u> 223, 661 (1984); S. Rodenhuis, <u>N.</u> Engl. J. Med. 317, 929 (1987); M. Barbacid, Eur. J. Clin. Invest. 20, 225 (1990); F. Sanger et al., J. Mol. Biol. 13, 373 (1975)), point mutations were screened for using RNAse protection assays (R.M. Myers et al., Science 230, 1242 (1985); E. Winter et al., Proc. Natl. Acad. Sci. USA 82, 7575 (1985)). Figure 4 shows a typical protection assay using a c-raf-1 probe. In this experiment the probe used covered the 3' end of raf-1 from the 3' most StuI site to the end of the coding 10 sequence. The first lane is a marker (pBR322 digested with HaeIII), the second shows the probe alone undigested, the third lane shows the probe hybridized to unrelated RNA in this case tRNA, the fourth lane shows hybridization with v-raf transformed cells and the lower 15 bands represent cleavage at points where the mouse c-<u>raf</u>-1 gene differs from v-<u>raf</u>. The fifth lane shows hybridization with RNA isolated from a normal lung of an untreated F1 mouse, the next lanes are RNA isolated from several tumors. In the case of the normal RNA, only 20 one, fully protected, band is detected while in the case of the tumors two major bands are seen after digestion. 20 out of 20 tumors analyzed in this fashion showed this extra band. These data demonstrate the following major points: 1) there is a tumor specific alteration in c-25 raf-1 that results in a region of non-homology recognizable by either RNAse A or T1; 2) The alterations are confined to the same region of one allele as two bands of equal size are present in the tumor lanes, and; 3) both alleles were expressed at comparable levels as 30 both bands are of approximately equal intensity. assay shown 5 µg of poly(A)+ RNA was hybridized from normal tissue, and 1 µg was used from the tumors. This was necessary to get signals that could be compared on 35 the same gel due to the overexpression of c-raf-1 in the tumors. By running these assays with various markers it was possible to estimate the approximate site of the

alteration(s) to be in the vicinity of the exon 14/exon 15 junction. In order to define the precise genetic alteration or alterations, PCR primers were designed which would generate a 600 bp fragment encompassing this region. cDNAs from tumor derived RNA were then 5 amplified and cloned into KS+ (Stratagene) for double stranded sequencing. The sequencing results from several tumors are shown in Figure 5. The top portion of Figure 5 presents a cartoon of the mouse Raf-1 protein. There are three conserved regions CR1, CR2 and CR3 with CR3 representing the kinase domain. The probe 10 used in the RNAse protection assays covers the indicated area, and the PCR primers amplified the bracketed region. Sequencing through this area revealed a variety of mutations just downstream of the APE site. These 15 mutants are shown in an expanded version at the bottom of Figure 5 (See also SEQ ID NO:1 for normal mouse sequence and SEQ ID NO:2, 3, 4, and 5 for mutant sequences). These mutants were isolated from four separate tumors, and in each case a normal allele (SEQ 20 ID NO:1) was also sequenced. Repeating the cDNA synthesis, PCR amplification, cloning and sequencing gives the same sequence and normal tissue shows no mutations demonstrating that these alterations are not artifactual. Sequence covering the amplified region has been examined and it is interesting that all of these 25 changes occur within a very small region of the raf protein. In fact the region where these mutations occur overlaps an epitope shared by monoclonal antibodies generated against raf (W. Kolch et al., Oncogene 5, 713 (1990)), and computer modeling of the protein shows this 30 to be a hydrophilic domain, the structure of which is predicted to be altered by these mutations. This indicates a biologically important region for the molecule and indeed the first of these mutation tested in NIH3T3 cell assays, after cloning into a retroviral 35 expression vector (E1-neo, (G. Heidecker et al., Mol.

WO 93/06248 PCT/US92/07817

-19-

Cell. Biol. 10, 2503 (1990))), was found to be weakly transforming when driven by a Moloney LTR. The transformation efficiency was comparable to EC2, a previously characterized mutation of human c-raf-1 cDNA (G. Heidecker et al., Mol. Cell. Biol. 10, 2503 (1990); C. Wasylyk et al., Mol. Cell. Biol. 9, 2247 (1989)) and ~20 fold lower than the v-raf oncogene.

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All publications mentioned hereinabove are hereby incorporated in their entirety by reference.

while the foregoing invention has been described in some detail for purposes of clarity and understanding, it will be appreciated by one skilled in the art from a reading of this disclosure that various changes in form and detail can be made without departing from the true scope of the invention and appended claims.

-20-

SEQUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: Rapp, Ulf R. Storm, Stephen M.
 - (ii) TITLE OF INVENTION: DETECTION METHOD FOR C-RAF-1 GENES
 - (iii) NUMBER OF SEQUENCES: 12
 - (iv) CORRESPONDENCE ADDRESS:

- (A) ADDRESSEE: NEEDLE & ROSENBERG, P.C. (B) STREET: Suite 400, 133 Carnegie Way, N.W.
- (C) CITY: Atlanta (D) STATE: Georgia
- (E) COUNTRY: USA
- (F) ZIP: 30303-1031
- (v) COMPUTER READABLE FORM:
- (A) MEDIUM TYPE: Floppy disk (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: (B) FILING DATE:

 - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:

 - (A) NAME: Perryman, David G. (B) REGISTRATION NUMBER: 33,438
 - (C) REFERENCE/DOCKET NUMBER: 1414.0421
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: (404) 688-0770
 - (B) TELEFAX: (404) 688-9880
- (2) INFORMATION FOR SEQ ID NO:1:
 - (1) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 648 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

Met Glu His Ile Gln Gly Ala Trp Lys Thr Ile Ser Asn Gly Phe Gly 5

Leu Lys Asp Ala Val Phe Asp Gly Ser Ser Cys Ile Ser Pro Thr Ile Val Gln Gln Phe Gly Tyr Gln Arg Arg Ala Ser Asp Asp Gly Lys Leu Thr Asp Ser Ser Lys Thr Ser Asn Thr Ile Arg Val Phe Leu Pro Asn Lys Gln Arg Thr Val Val Asn Val Arg Asn Gly Met Ser Leu His Asp Cys Leu Met Lys Ala Leu Lys Val Arg Gly Leu Gin Pro Glu Cys Cys Ala Val Phe Arg Leu Leu Gln Glu His Lys Gly Lys Lys Ala Arg Leu Asp Trp Asn Thr Asp Ala Ala Ser Leu Ile Gly Glu Glu Leu Gln Val 120 Asp Phe Leu Asp His Val Pro Ile Thr Thr His Asn Phe Ala Arg Lys Thr Phe Leu Lys Leu Ala Phe Cys Asp Ile Cys Gln Lys Phe Leu Leu Asn Gly Phe Arg Cys Gln Thr Cys Gly Tyr Lys Phe His Glu His Cys Ser Thr Lys Val Pro Thr Met Cys Val Asp Trp Ser Asn Ile Arg Gln Leu Leu Leu Phe Pro Asn Ser Thr Val Gly Asp Ser Gly Val Pro Ala 205 200 195 Pro Pro Ser Phe Pro Met Arg Arg Met Arg Glu Ser Val Ser Arg Met Pro Ala Ser Ser Gln His Arg Tyr Ser Thr Pro His Ala Phe Thr Phe 235 Asn Thr Ser Ser Pro Ser Ser Glu Gly Ser Leu Ser Gln Arg Gln Arg Ser Thr Ser Thr Pro Asn Val His Met Val Ser Thr Thr Leu His Val 265 Asp Ser Arg Met Ile Glu Asp Ala Ile Arg Ser His Ser Glu Ser Ala 280 Ser Pro Ser Ala Leu Ser Ser Ser Pro Asn Asn Leu Ser Pro Thr Gly 295

305					210				Ala	-					
G1y	Ser	Gly	Thr	G1n 325	G1n	Lys	Asn	Lys	Ile 330	Arg	Pro	Arg	G1y	G1n 335	Arg
Asp	Ser	Ser	Tyr 340	Tyr	Trp	Glu	Ile	G1u 345	Ala	Ser	Glu	Val	Met 350	Leu	Ser
		355					200		Thr						
	370					3/3			Lys						
385		-			390				Val						
				405					Gly 410						
Leu	Ala	Ile	Va1 420	Thr	Gln	Trp	Cys	G1u 425	Gly	Ser	Ser	Leu	Tyr 430	Lys	His
		435					440								Ile
	450					455					•				Ile
465					4/0					•• -					Leu 480
				485											Trp
			500					500							Met
		515					520	1							Gln
	530					J_J_	,								Gly
545	Leu	Pro			220	'									Met 560
Val	Gly			505)				•••						Asn
Cys	Pro	Lys	Ala 580	Met	Lys	Arg	, Lei	Val 585	Ala S	. Asp	Cys	Va1	Lys 590	Lys	s Val

Lys Glu Glu Arg Pro Leu Phe Pro Gln Ile Leu Ser Ser Ile Glu Leu 595 600 605

Leu Gln His Ser Leu Pro Lys Ile Asn Arg Ser Ala Ser Glu Pro Ser 610 615 620

Leu His Arg Ala Ala His Thr Glu Asp Ile Asn Ala Cys Thr Leu Thr 625 630 640

Thr Ser Pro Arg Leu Pro Val Phe 645

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 648 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Glu His Ile Gln Gly Ala Trp Lys Thr Ile Ser Asn Gly Phe Gly
1 10 15

Leu Lys Asp Ala Val Phe Asp Gly Ser Ser Cys Ile Ser Pro Thr Ile 20 25

Val Gln Gln Phe Gly Tyr Gln Arg Arg Ala Ser Asp Asp Gly Lys Leu 35 40 45

Thr Asp Ser Ser Lys Thr Ser Asn Thr Ile Arg Val Phe Leu Pro Asn 50 60

Lys Gln Arg Thr Val Val Asn Val Arg Asn Gly Met Ser Leu His Asp 65 70 80

Cys Leu Met Lys Ala Leu Lys Val Arg Gly Leu Gln Pro Glu Cys Cys 85 90 95

Ala Val Phe Arg Leu Leu Gln Glu His Lys Gly Lys Lys Ala Arg Leu 100 105 110

Asp Trp Asn Thr Asp Ala Ala Ser Leu Ile Gly Glu Glu Leu Gln Val

Asp Phe Leu Asp His Val Pro Ile Thr Thr His Asn Phe Ala Arg Lys 130 135 140

Thr Phe Leu Lys Leu Ala Phe Cys Asp Ile Cys Gln Lys Phe Leu Leu 145 150 160

Asn Gly Phe Arg Cys Gln Thr Cys Gly Tyr Lys Phe His Glu His Cys 175

Ser Thr Lys Val Pro Thr Met Cys Val Asp Trp Ser Asn Ile Arg Gln 180 Leu Leu Leu Phe Pro Asn Ser Thr Val Gly Asp Ser Gly Val Pro Ala Pro Pro Ser Phe Pro Met Arg Arg Met Arg Glu Ser Val Ser Arg Met Pro Ala Ser Ser Gln His Arg Tyr Ser Thr Pro His Ala Phe Thr Phe 230 Asn Thr Ser Ser Pro Ser Ser Glu Gly Ser Leu Ser Gln Arg Gln Arg 250 Ser Thr Ser Thr Pro Asn Val His Met Val Ser Thr Thr Leu His Val Asp Ser Arg Met Ile Glu Asp Ala Ile Arg Ser His Ser Glu Ser Ala 285 Ser Pro Ser Ala Leu Ser Ser Ser Pro Asn Asn Leu Ser Pro Thr Gly Trp Ser Gln Pro Lys Thr Pro Val Pro Ala Gln Arg Glu Arg Ala Pro Gly Ser Gly Thr Gln Gln Lys Asn Lys Ile Arg Pro Arg Gly Gln Arg Asp Ser Ser Tyr Tyr Trp Glu Ile Glu Ala Ser Glu Val Met Leu Ser 345 Thr Arg Ile Gly Ser Gly Ser Phe Gly Thr Val Tyr Lys Gly Lys Trp 360 His Gly Asp Val Ala Val Lys Ile Leu Lys Val Val Asp Pro Thr Pro 370 380 Glu Gin Leu Gin Ala Phe Arg Asn Glu Val Ala Val Leu Arg Lys Thr 390 Arg His Val Asn Ile Leu Leu Phe Met Gly Tyr Met Thr Lys Asp Asn 405 Leu Ala Ile Val Thr Gln Trp Cys Glu Gly Ser Ser Leu Tyr Lys His Leu His Val Gln Glu Thr Lys Phe Gln Met Phe Gln Leu Ile Asp Ile Ala Arg Gln Thr Ala Gln Gly Met Asp Tyr Leu His Ala Lys Asn Ile 455

Ile His Arg Asp Met Lys Ser Asn Asn Ile Phe Leu His Glu Gly Leu 465 Thr Val Lys Ile Gly Asp Phe Gly Leu Ala Thr Val Lys Ser Arg Trp Ser Gly Ser Gln Gln Val Glu Gln Pro Thr Gly Ser Val Leu Trp Met 505 500 Ala Pro Glu Val Val Arg Met Gln Asp Asp Asn Pro Phe Ser Phe Gln Ser Asp Val Tyr Ser Tyr Gly Ile Val Leu Tyr Glu Leu Met Ala Gly Glu Leu Pro Tyr Ala His Ile Asn Asn Arg Asp Gln Ile Ile Phe Met Val Gly Arg Gly Tyr Ala Ser Pro Asp Leu Ser Arg Leu Tyr Lys Asn Cys Pro Lys Ala Met Lys Arg Leu Val Ala Asp Cys Val Lys Lys Val Lys Glu Glu Arg Pro Leu Phe Pro Gln Ile Leu Ser Ser Ile Glu Leu Leu Gln His Ser Leu Pro Lys Ile Asn Arg Ser Ala Ser Glu Pro Ser Leu His Arg Ala Ala His Thr Glu Asp Ile Asn Ala Cys Thr Leu Thr 630 Thr Ser Pro Arg Leu Pro Val Phe 645

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 648 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Met Glu His Ile Gln Gly Ala Trp Lys Thr Ile Ser Asn Gly Phe Gly 10 15

Leu Lys Asp Ala Val Phe Asp Gly Ser Ser Cys Ile Ser Pro Thr Ile 20 25 30

Val Gln Gln Phe Gly Tyr Gln Arg Arg Ala Ser Asp Asp Gly Lys Leu

	Asp 50					23					• •				
65	G1 n				/0					• -					
-	Leu			83											
	Val		100					100							
	Trp	115					120								
	130					133				•					Lys
145					120										Leu 160
				Too											Cys
			180					100							Gln
		195					200								Ala
	210					213					-				Met
225					230										Phe 240
				245											Arg
			260					LUZ	,						: Val
		2/5)												· Ala
	290)				295	,				-				- Gly
305	5				210	,									320
G1)	, Set	· G1)	/ Thr	• G1r 325	Glr	Lys	s Asr	ı Ly:	330	e Arg	g Pro	Arg	g G1)	/ Gl:	n Arg

Asp Ser Ser Tyr Tyr Trp Glu Ile Glu Ala Ser Glu Val Met Leu Ser 345 Thr Arg Ile Gly Ser Gly Ser Phe Gly Thr Val Tyr Lys Gly Lys Trp 355 360 365 His Gly Asp Val Ala Val Lys Ile Leu Lys Val Val Asp Pro Thr Pro Glu Gln Leu Gln Ala Phe Arg Asn Glu Val Ala Val Leu Arg Lys Thr 395 Arg His Val Asn Ile Leu Leu Phe Met Gly Tyr Met Thr Lys Asp Asn Leu Ala Ile Val Thr Gln Trp Cys Glu Gly Ser Ser Leu Tyr Lys His Leu His Val Gln Glu Thr Lys Phe Gln Met Phe Gln Leu Ile Asp Ile Ala Arg Gln Thr Ala Gln Gly Met Asp Tyr Leu His Ala Lys Asn Ile Ile His Arg Asp Met Lys Ser Asn Asn Ile Phe Leu His Glu Gly Leu Thr Val Lys Ite Gly Asp Phe Gly Leu Ala Thr Val Lys Ser Arg Trp Ser Gly Ser Gln Gln Val Glu Gln Pro Thr Gly Ser Val Leu Trp Met 505 Ala Pro Glu Val Ile Arg Met Gln Asp Asn Asn Pro Phe Ser Phe Gln 520 Ser Asp Val Tyr Ser Tyr Gly Ile Val Leu Tyr Glu Leu Met Ala Gly

Glu Leu Pro Tyr Ala His Ile Asn Asn Arg Asp Gln Ile Ile Phe Met 545

Val Gly Arg Gly Tyr Ala Ser Pro Asp Leu Ser Arg Leu Tyr Lys Asn

Cys Pro Lys Ala Met Lys Arg Leu Val Ala Asp Cys Val Lys Lys Val 580 585 590

Lys Glu Glu Arg Pro Leu Phe Pro Gln Ile Leu Ser Ser Ile Glu Leu 595 600 605

Leu Gln His Ser Leu Pro Lys Ile Asn Arg Ser Ala Ser Glu Pro Ser 610 620 Leu His Arg Ala Ala His Thr Glu Asp Ile Asn Ala Cys Thr Leu Thr 625

Thr Ser Pro Arg Leu Pro Val Phe

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 648 amino acids

645

- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Glu His IIe Gln Gly Ala Trp Lys Thr IIe Ser Asn Gly Phe Gly 15

Leu Lys Asp Ala Val Phe Asp Gly Ser Ser Cys Ile Ser Pro Thr Ile 20

Val Gln Gln Phe Gly Tyr Gln Arg Arg Ala Ser Asp Asp Gly Lys Leu 35 40 45

Thr Asp Ser Ser Lys Thr Ser Asn Thr Ile Arg Val Phe Leu Pro Asn 50 60

Lys Gln Arg Thr Val Val Asn Val Arg Asn Gly Met Ser Leu His Asp 80

Cys Leu Met Lys Ala Leu Lys Val Arg Gly Leu Gln Pro Glu Cys Cys

Ala Val Phe Arg Leu Leu Gln Glu His Lys Gly Lys Lys Ala Arg Leu 100 105 110

Asp Trp Asn Thr Asp Ala Ala Ser Leu Ile Gly Glu Glu Leu Gln Val

Asp Phe Leu Asp His Val Pro Ile Thr Thr His Asn Phe Ala Arg Lys
130 135

Thr Phe Leu Lys Leu Ala Phe Cys Asp Ile Cys Gln Lys Phe Leu Leu 150 155 160

Asn Gly Phe Arg Cys Gln Thr Cys Gly Tyr Lys Phe His Glu His Cys 175

Ser Thr Lys Val Pro Thr Met Cys Val Asp Trp Ser Asn Ile Arg Gln
180 185

Leu Leu Leu Phe Pro Asn Ser Thr Val Gly Asp Ser Gly Val Pro Ala 200

Pro	Pro 210	Ser	Phe	Pro	Met	Arg 215	Arg	Met	Arg	Glu	Ser 220	Val	Ser	Arg	Met
Pro 225	Ala	Ser	Ser	Gln	His 230	Arg	Tyr	Ser	Thr	Pro 235	His	Ala	Phe	Thr	Phe 240
Asn	Thr	Ser	Ser	Pro 245	Ser	Ser	Glu	Gly ·	Ser 250	Leu	Ser	Gln	Arg.	G1n 255	Arg
Ser	Thr	Ser	Thr 260	Pro	Asn	Va1	His	Met 265	Val	Ser	Thr	Thr	Leu 270	His	Val
Asp	Ser	Arg 275	Met	Ile	Glu	Asp	A1 a 280	Ile	Arg	Ser	His	Ser 285	Glu	Ser	Ala
Ser	Pro 290	Ser	Ala	Leu	Ser	Ser 295	Ser	Pro	Asn	Asn	Leu 300	Ser	Pro	Thr	G1y
Trp 305	Ser	Gln	Pro	Lys	Thr 310	Pro	Val	Pro	Ala	G1n 315	Arg	Glu	Arg	Ala	Pro 320
G1 y	Ser	Gly	Thr	G1n 325	Gln	Lys	Asn	Lys	11e. 330	Arg	Pro	Arg	Gly	G1n 335	Arg
Asp	Ser	Ser	Tyr 340	Tyr	Trp	Glu	Ile	G1u 345	Ala	Ser	Glu	Val	Met 350	Leu	Ser
Thr	Arg	Ile 355	Gly	Ser	Gly	Ser	Phe 360	Gly	Thr	Val	Tyr	Lys 365	Gly	Lys	Trp
	370					375				•	Va1 380				
G1 u 385	Gln	Leu	Gln	Ala	Phe 390	Arg	Asn	Glu	Val	A1a 395	Val	Leu	Arg	Lys	Thr 400
Arg	His	Val	Asn	Ile 405	Leu	Leu	Phe	Met	Gly 410	Tyr	Met	Thr	Lys	Asp 415	Asr
			420					425			Ser		430		
Leu	His	Va1 435	Gln	Gļu	Thr	Lys	Phe 440	Gln	Met	Phe	Gln	Leu 445	Ile	Asp	116
Ala	Arg 450	Gln	Thr	Ala	Gln	G1y 455	Met	Asp	Tyr	Leu	His 460	Ala	Lys	Asn	Ile
.I1e 465		Arg	Asp	Met	Lys 470	Ser	Asn	Asn	Ile	Phe 475	Leu	His	Glu	Gly	Le:
Thr	Val	Lys	Ile	Gly	Asp	Phe	Gly	Leu	A1a 490	Thr	Val	Lys	Ser	Arg 495	Tr

 Ser Gly Ser
 Gln Gln Gln Val
 Glu Gln Pro 505
 Thr Gly Ser Val
 Leu Trp Met 510

 Ala Pro Glu Val
 Ile Arg Met 520
 Asp Asp Asn Pro 525
 Ser Ser Gln 525

 Ser Asp Val
 Tyr Ser Tyr Gly 11e Val
 Leu Tyr Glu Leu Met Ala Gly 535

 Glu Leu Pro Tyr Ala His 550
 Ile Asn Asn Arg Asp Gln Ile Ile Phe Met 560

 Val Gly Arg Gly Tyr Ala Ser Pro Asp Leu 555
 Ser Arg Leu Tyr Lys Asn 575

 Cys Pro Lys Ala Met Lys Arg Leu 585
 Ala Asp Cys Val Lys Lys Val 590

 Lys Glu Glu Arg Pro Leu Phe Pro 600
 Gln Ile Leu Ser Ser Ile Glu Leu 605

 Leu Gln His Ser Leu Pro Lys 615
 Ile Asn Arg Ser Ala Ser Glu Pro Ser 620

Leu His Arg Ala Ala His Thr Glu Asp Ile Asn Ala Cys Thr Leu Thr 625 630 640

Thr Ser Pro Arg Leu Pro Val Phe 645

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 648 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Met Glu His Ile Gln Gly Ala Trp Lys Thr Ile Ser Asn Gly Phe Gly
10 15

Leu Lys Asp Ala Val Phe Asp Gly Ser Ser Cys Ile Ser Pro Thr Ile 20 30

Val Gln Gln Phe Gly Tyr Gln Arg Arg Ala Ser Asp Asp Gly Lys Leu 35 40 45

Thr Asp Ser Ser Lys Thr Ser Asn Thr Ile Arg Val Phe Leu Pro Asn 50

Lys Gln Arg Thr Val Val Asn Val Arg Asn Gly Met Ser Leu His Asp 80

Cys Leu Met Lys Ala Leu Lys Val Arg Gly Leu Gln Pro Glu Cys Cys Ala Val Phe Arg Leu Leu Gln Glu His Lys Gly Lys Lys Ala Arg Leu 105 Asp Trp Asn Thr Asp Ala Ala Ser Leu Ile Gly Glu Glu Leu Gln Val Asp Phe Leu Asp His Val Pro Ile Thr Thr His Asn Phe Ala Arg Lys Thr Phe Leu Lys Leu Ala Phe Cys Asp Ile Cys Gln Lys Phe Leu Leu Asn Gly Phe Arg Cys Gln Thr Cys Gly Tyr Lys Phe His Glu His Cys Ser Thr Lys Val Pro Thr Met Cys Val Asp Trp Ser Asn Ile Arg Gln Leu Leu Leu Phe Pro Asn Ser Thr Val Gly Asp Ser Gly Val Pro Ala 200 Pro Pro Ser Phe Pro Met Arg Arg Met Arg Glu Ser Val Ser Arg Met Pro Ala Ser Ser Gln His Arg Tyr Ser Thr Pro His Ala Phe Thr Phe 235 Asn Thr Ser Ser Pro Ser Ser Glu Gly Ser Leu Ser Gln Arg Gln Arg 250 Ser Thr Ser Thr Pro Asn Val His Met Val Ser Thr Thr Leu His Val 265 Asp Ser Arg Met Ile Glu Asp Ala Ile Arg Ser His Ser Glu Ser Ala 280 Ser Pro Ser Ala Leu Ser Ser Ser Pro Asn Asn Leu Ser Pro Thr Gly 295 Trp Ser Gin Pro Lys Thr Pro Val Pro Ala Gin Arg Glu Arg Ala Pro Gly Ser Gly Thr Gln Gln Lys Asn Lys Ile Arg Pro Arg Gly Gln Arg Asp Ser Ser Tyr Tyr Trp Glu Ile Glu Ala Ser Glu Val Met Leu Ser 345 Thr Arg Ile Gly Ser Gly Ser Phe Gly Thr Val Tyr Lys Gly Lys Trp 360

His Gly Asp Val Ala Val Lys Ile Leu Lys Val Val Asp Pro Thr Pro Glu Gln Leu Gln Ala Phe Arg Asn Glu Val Ala Val Leu Arg Lys Thr Arg His Val Asn Ile Leu Leu Phe Met Gly Tyr Met Thr Lys Asp Asn Leu Ala Ile Val Thr Gln Trp Cys Glu Gly Ser Ser Leu Tyr Lys His Leu His Val Gln Glu Thr Lys Phe Gln Met Phe Gln Leu Ile Asp Ile Ala Arg Gin Thr Ala Gin Gly Met Asp Tyr Leu His Ala Lys Asn Ile Ile His Arg Asp Met Lys Ser Asn Asn Ile Phe Leu His Glu Gly Leu Thr Val Lys Ile Gly Asp Phe Gly Leu Ala Thr Val Lys Ser Arg Trp 490 Ser Gly Ser Gln Gln Val Glu Gln Pro Thr Gly Ser Val Leu Trp Met 505 Ala Pro Glu Val Ile Arg Met Gln Asp Asp Asn Pro Phe Ser Phe Gln Ser Thr Cys Thr Phe Tyr Gly Ile Val Leu Tyr Glu Leu Met Ala Gly 530 Glu Leu Pro Tyr Ala His Ile Asn Asn Arg Asp Gln Ile Ile Phe Met Val Gly Arg Gly Tyr Ala Ser Pro Asp Leu Ser Arg Leu Tyr Lys Asn 570 Cys Pro Lys Ala Met Lys Arg Leu Val Ala Asp Cys Val Lys Lys Val 585 Lys Glu Glu Arg Pro Leu Phe Pro Gln Ile Leu Ser Ser Ile Glu Leu Leu Gln His Ser Leu Pro Lys Ile Asn Arg Ser Ala Ser Glu Pro Ser Leu His Arg Ala Ala His Thr Glu Asp Ile Asn Ala Cys Thr Leu Thr 630 625 Thr Ser Pro Arg Leu Pro Val Phe 645

(2) INFORMATION FOR SEQ ID NO:6:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:	
AACTTGTGGT GGTTGGACCT	20
(2) INFORMATION FOR SEQ ID NO:7:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:	
TCATCCACAA AGTGATTCTG	20
(2) INFORMATION FOR SEQ ID NO:8:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(x1) SEQUENCE DESCRIPTION: SEQ ID NO:8:	
AGGAGACCAA GTTTCAGATG	20
(2) INFORMATION FOR SEQ ID NO:9:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:	
GCGTGCAAGC ATTGATATCC	20
(2) INFORMATION FOR SEQ ID NO:10:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1947 base pairs	

(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

	Edoruge 222		-			
				GCTTTGGACT		60
				AGCAGTTTGG		120
				CAAGCAATAC		180
TTCTTGCCGA	ATAAGCAAAG	GACTGTGGTC	AATGTGCGGA	ATGGAATGAG	CTTACATGAC	240
				AGTGCTGTGC		300
				GGAACACCGA		360
				TTCCCATCAC		420
TTTGCTCGGA	AAACGTTCCT	GAAGCTTGCA	TTCTGTGACA	TCTGTCAGAA	GTTCCTGCTA	480
				AGCACTGTAG		540
CCTACTATGT	GTGTGGACTG	GAGTAATATC	AGACAGCTCT	TGCTGTTTCC	AAATTCCACT	600
GTTGGTGACA	GTGGAGTCCC	AGCACCACCT	TCTTTCCCAA	TGCGTCGGAT	GCGAGAATCT	660
GTTTCCCGGA	TGCCTGCTAG	TTCCCAGCAC	AGATACTCTA	CACCCCATGC	CTTCACTTTC	720
AACACCTCCA	GCCCTTCCTC	AGAAGGTTCC	CTCTCCCAGA	GGCAGAGGTC	AACGTCCACT	780
CCCAATGTCC	ACATGGTCAG	CACCACCCTG	CATGTGGACA	GCAGGATGAT	TGAGGATGCA	840
ATTCGAAGTC	ACAGTGAATC	AGCCTCACCT	TCAGCCCTGT	CCAGCAGCCC	AAACAACCTG	900
GGTCCAACAG	GCTGGTCACA	GCCCAAAACC	CCCGTGCCAG	CACAAAGAGA	GCGGGCACCA	960
GGATCTGGGA	CCCAGCAAAA	AAACAAAATT	AGGCCTCGTG	GGCAGAGAGA	CTCGAGTTAT	1020
TACTGGGAAA	TAGAAGCCAG	TGAGGTGATG	CTGTCTACTC	GGATCGGGTC	AGGTTCCTTT	1080
GGCACTGTGT	ACAAGGGCAA	GTGGCATGGA	GATGTTGCAG	TAAAGATCCT	AAAGGTGGTT	1140
GACCCAACTC	CAGAGCAACT	TCAGGCCTTC	AGGAACGAGG	TGGCTGTTTT	GCGCAAAACA	1200
CGGCATGTTA	ACATCCTGCT	GTTCATGGGG	TACATGACAA	AGGACAACCT	GGCGATTGTG	1260
ACTCAGTGGT	GTGAAGGCAG	CAGTCTCTAC	AAACACCTGC	ATGTCCAGGA	GACCAAATTC	1320
CAGATGTTCC	AGCTAATTGA	CATTGCCCGA	CAGACAGCTC	AGGGAATGGA	CTATTTGCAT	1380
GCAAAGAACA	TCATCCACAG	AGACATGAAA	TCCAACAATA	TATTTCTCCA	TGAAGGCCTC	1440
ACGGTGAAAA	TTGGAGATTT	TGGTTTGGCA	ACAGTGAAGT	CACGCTGGAG	TGGTTCTCAG	1500
CAGGTTGAAC	AGCCCACTGG	CTCTGTGCTG	TGGATGGCCC	CAGAAGTAAT	CCGGATGCAG	1560

GATGACAACC CGTTCAGCTT CCAGTCCGAC GTGTACTCGT ACGGCATCGT GCTGTACGAG 1620
CTGATGGCTG GGGAGCTTCC CTACGCCCAC ATCAACAACC GAGACCAGAT CATCTTCATG 1680
GTAGGCCGTG GGTATGCATC CCCTGATCTC AGCAGGCTCT ACAAGAACTG CCCCAAGGCA 1740
ATGAAGAGGT TGGTGGCTGA CTGTGTGAAG AAAGTCAAAG AAGAGAGACC TTTGTTTCCC 1800
CAGATCCTGT CTTCCATCGA GCTGCTTCAG CACTCTCTGC CGAAAATCAA CAGGAGCGCC 1860
TCTGAGCCTT CCCTGCATCG GGCAGCTCAC ACTGAGGACA TCAATGCTTG CACGCTGACT 1920
ACATCCCCAA GGCTACCAGT CTTCTAG 1947

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1947 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1..1944
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:
- ATG GAG CAC ATA CAG GGA GCT TGG AAG ACG ATC AGC AAT GGT TTT GGA

 Met Glu His Ile Gln Gly Ala Trp Lys Thr Ile Ser Asn Gly Phe Gly
- TTC AAA GAT GCC GTG TTT GAT GGC TCC AGC TGC ATC TCT CCT ACA ATA 96
 Phe Lys Asp Ala Val Phe Asp Gly Ser Ser Cys Ile Ser Pro Thr Ile
- GTT CAG CAG TTT GGC TAT CAG CGC CGG GCA TCA GAT GAT GGC AAA CTC 144

 Val Gln Gln Phe Gly Tyr Gln Arg Arg Ala Ser Asp Asp Gly Lys Leu
 45
- ACA GAT CCT TCT AAG ACA AGC AAC ACT ATC CGT GTT TTC TTG CCG AAC 192
 Thr Asp Pro Ser Lys Thr Ser Asn Thr Ile Arg Val Phe Leu Pro Asn
 50
 60
- AAG CAA AGA ACA GTG GTC AAT GTG CGA AAT GGA ATG AGC TTG CAT GAC 240

 Lys Gln Arg Thr Val Val Asn Val Arg Asn Gly Met Ser Leu His Asp
 65 70 75 80

TGC	стт	ATG	AAA	GCA	CTC	AAG	GTG	AGG	GGC	CTG	CAA	CCA	GAG	TGC	TGT	288
Cys	Leu	Met	Ĺys	A1a 85	Leu	Lys	Val	Arg	G1 <i>y</i> 90	Leu	Gln	Pro	Glu	Cys 95	Cys	
GCA	GTG.	TTC	AGA	CTT	стс	CAC	GAA	CAC	AAA	GGT	AAA	AAA	GCA	CGC	TTA	336
Ala	Val	Phe	Arg 100	Leu	Leu	His	Glu	His 105	Lys	Gly	Lys	Lys	Ala 110	Arg	Leu	
GAT	TGG	AAT	ACT	GAT	GCT	GCG	TCT	TTG	ATT	GGA	GAA	GAA	CTT	CAA	GTA	384
		115					120				Glu					
GAT	TTC	CTG	GAT	CAT	GTT	CCC	CTC	ACA	ACA	CAC	AAC	ш	GCT	CGG	AAG	432
	130					135					Asn 140					
											CAG					480
145					150					133	Gln					
											Ш					528
	•			165					170		Phe					
															CAA	576
			180					193			Ser		170			
															GCA	624
		195					200				Ser					
											TCT					672
	210					210					Ser 220					
ССТ	GTT	AGT	TCT	CAG	CAC	AGA	TAT	TCT	ACA	CCT	CAC	GCC	TTC	ACC	TTT	720
225					230					233	His					
															AGG	768
				245					250		Ser					
TCG	ACA	TCC	ACA	ССТ	AAT	GTC	CAC	ATG	GTC	AGC	ACC	ACG	CTG	CCT	GTG	816

Ser Thr Ser Thr Pro Asn Val His Met Val Ser Thr Thr Leu Pro Val 265 260 GAC AGC AGG ATG ATT GAG GAT GCA ATT CGA AGT CAC AGC GAA TCA GCC Asp Ser Arg Met Ile Glu Asp Ala Ile Arg Ser His Ser Glu Ser Ala TCA CCT TCA GCC CTG TCC AGT AGC CCC AAC AAT CTG AGC CCA ACA GGC 912 Ser Pro Ser Ala Leu Ser Ser Ser Pro Asn Asn Leu Ser Pro Thr Gly 290 TGG TCA CAG CCG AAA ACC CCC GTG CCA GCA CAA AGA GAG CGG GCA CCA 960 Trp Ser Gln Pro Lys Thr Pro Val Pro Ala Gln Arg Glu Arg Ala Pro GTA TCT GGG ACC CAG GAG AAA AAC AAA ATT AGG CCT CGT GGA CAG AGA 1008 Val Ser Gly Thr Gln Glu Lys Asn Lys Ile Arg Pro Arg Gly Gln Arg GAT TCA AGC TAT TAT TGG GAA ATA GAA GCC AGT GAA GTG ATG CTG TCC 1056 Asp Ser Ser Tyr Tyr Trp Glu Ile Glu Ala Ser Glu Val Met Leu Ser ACT CGG ATT GGG TCA GGC TCT TTT GGA ACT GTT TAT AAG GGT AAA TGG 1104 Thr Arg Ile Gly Ser Gly Ser Phe Gly Thr Val Tyr Lys Gly Lys Trp 355 CAC GGA GAT GTT GCA GTA AAG ATC CTA AAG GTT GTC GAC CCA ACC CCA 1152 His Gly Asp Val Ala Val Lys Ile Leu Lys Val Val Asp Pro Thr Pro GAG CAA TTC CAG GCC TTC AGG AAT GAG GTG GCT GTT CTG CGC AAA ACA 1200 Glu Gln Phe Gln Ala Phe Arg Asn Glu Val Ala Val Leu Arg Lys Thr 385 CGG CAT GTG AAC ATT CTG CTT TTC ATG GGG TAC ATG ACA AAG GAC AAC 1248 Arg His Val Asn Ile Leu Leu Phe Met Gly Tyr Met Thr Lys Asp Asn CTG GCA ATT GTG ACC CAG TGG TGC GAG GGC AGC CTC TAC AAA CAC 1296 Leu Ala Ile Val Thr Gln Trp Cys Glu Gly Ser Ser Leu Tyr Lys His 420 CTG CAT GTC CAG GAG ACC AAG TTT CAG ATG TTC CAG CTA ATT GAC ATT 1344 Leu His Val Gln Glu Thr Lys Phe Gln Met Phe Gln Leu Ile Asp Ile 440 435 GCC EGG CAG ACG GCT CAG GGA ATG GAC TAT TTG CAT GCA AAG AAC ATC 1392 Ala Arg Gin Thr Ala Gin Gly Met Asp Tyr Leu His Ala Lys Asn Ile ATC CAT AGA GAC ATG AAA TCC AAC AAT ATA TTT CTC CAT GAA GGC TTA 1440 Ile His Arg Asp Met Lys Ser Asn Asn Ile Phe Leu His Glu Gly Leu ACA GTG AAA ATT GGA GAT TTT GGT TTG GCA ACA GTA AAG TCA CGC TGG 1488 Thr Val Lys Ile Gly Asp Phe Gly Leu Ala Thr Val Lys Ser Arg Trp AGT GGT TCT CAG CAG GTT GAA CAA CCT ACT GGC TCT GTC CTC TGG ATG 1536 Ser Gly Ser Gln Gln Val Glu Gln Pro Thr Gly Ser Val Leu Trp Met 500 GCC CCA GAG GTG ATC CGA ATG CAG GAT AAC AAC CCA TTC AGT TTC CAG 1584 Ala Pro Glu Val Ile Arg Met Gln Asp Asn Asn Pro Phe Ser Phe Gln TCG GAT GTC TAC TCC TAT GGC ATC GTA TTG TAT GAA CTG ATG ACG GGG 1632 Ser Asp Val Tyr Ser Tyr Gly Ile Val Leu Tyr Glu Leu Met Thr Gly 535 530 GAG CTT CCT TAT TCT CAC ATC AAC CGA GAT CAG ATC ATC TTC ATG 1680 Glu Leu Pro Tyr Ser His Ile Asn Asn Arg Asp Gln Ile Ile Phe Met 545 GTG GGC CGA GGA TAT GCC TCC CCA GAT CTT AGT AAG CTA TAT AAG AAC 1728 Val Gly Arg Gly Tyr Ala Ser Pro Asp Leu Ser Lys Leu Tyr Lys Asn 565 TGC CCC AAA GCA ATG AAG AGG CTG GTA GCT GAC TGT GTG AAG AAA GTA 1776 Cys Pro Lys Ala Met Lys Arg Leu Val Ala Asp Cys Val Lys Lys Val AAG GAA GAG AGG CCT CTT TTT CCC CAG ATC CTG TCT TCC ATT GAG CTG 1824 Lys Glu Glu Arg Pro Leu Phe Pro Gln Ile Leu Ser Ser Ile Glu Leu 600 595

CTC CAA CAC TCT CTA CCG AAG ATC AAC CGG AGC GCT TCC GAG CCA TCC 1872

Leu Gln His Ser Leu Pro Lys Ile Asn Arg Ser Ala Ser Glu Pro Ser

610 615 620

TTG CAT CGG GCA GCC CAC ACT GAG GAT ATC AAT GCT TGC ACG CTG ACC 1920

Leu His Arg Ala Ala His Thr Glu Asp Ile Asn Ala Cys Thr Leu Thr 625 630 635 640

ACG TCC CCG AGG CTG CCT GTC TTC TAG

1947

Thr Ser Pro Arg Leu Pro Val Phe

- (2) INFORMATION FOR SEQ ID NO:12:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 648 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Met Glu His Ile Gln Gly Ala Trp Lys Thr Ile Ser Asn Gly Phe Gly
10 15

Phe Lys Asp Ala Val Phe Asp Gly Ser Ser Cys Ile Ser Pro Thr Ile 25 30

Val Gln Gln Phe Gly Tyr Gln Arg Arg Ala Ser Asp Asp Gly Lys Leu 35 40

Thr Asp Pro Ser Lys Thr Ser Asn Thr Ile Arg Val Phe Leu Pro Asn 50 55

Lys Gln Arg Thr Val Val Asn Val Arg Asn Gly Met Ser Leu His Asp
65 70 75 80

Cys Leu Met Lys Ala Leu Lys Val Arg Gly Leu Gln Pro Glu Cys Cys 90 95

Ala Val Phe Arg Leu Leu His Glu His Lys Gly Lys Lys Ala Arg Leu 100 105 110

Asp Trp Asn Thr Asp Ala Ala Ser Leu Ile Gly Glu Glu Leu Gln Val

Asp Phe Leu Asp His Val Pro Leu Thr Thr His Asn Phe Ala Arg Lys 130 135

Thr Phe Leu Lys Leu Ala Phe Cys Asp Ile Cys Gln Lys Phe Leu Leu 145 150 155 160

Asn Gly Phe Arg Cys Gln Thr Cys Gly Tyr Lys Phe His Glu His Cys 175

450

Ser Thr Lys Val Pro Thr Met Cys Val Asp Trp Ser Asn Ile Arg Gln Leu Leu Leu Phe Pro Asn Ser Thr Ile Gly Asp Ser Gly Val Pro Ala 200 Leu Pro Ser Leu Thr Met Arg Arg Met Arg Glu Ser Val Ser Arg Met Pro Val Ser Ser Gln His Arg Tyr Ser Thr Pro His Ala Phe Thr Phe Asn Thr Ser Ser Pro Ser Ser Glu Gly Ser Leu Ser Gln Arg Gln Arg 250 Ser Thr Ser Thr Pro Asn Val His Met Val Ser Thr Thr Leu Pro Val 265 Asp Ser Arg Met Ile Glu Asp Ala Ile Arg Ser His Ser Glu Ser Ala Ser Pro Ser Ala Leu Ser Ser Ser Pro Asn Asn Leu Ser Pro Thr Gly 300 Trp Ser Gln Pro Lys Thr Pro Val Pro Ala Gln Arg Glu Arg Ala Pro Val Ser Gly Thr Gln Glu Lys Asn Lys Ile Arg Pro Arg Gly Gln Arg 330 325 Asp Ser Ser Tyr Tyr Trp Glu Ile Glu Ala Ser Glu Val Met Leu Ser 345 Thr Arg Ile Gly Ser Gly Ser Phe Gly Thr Val Tyr Lys Gly Lys Trp 355 . 360 His Gly Asp Val Ala Val Lys Ile Leu Lys Val Val Asp Pro Thr Pro Glu Gin Phe Gin Ala Phe Arg Asn Glu Val Ala Val Leu Arg Lys Thr 395 385 Arg His Val Asn Ile Leu Leu Phe Met Gly Tyr Met Thr Lys Asp Asn 410 Leu Ala Ile Val Thr Gln Trp Cys Glu Gly Ser Ser Leu Tyr Lys His 420 Leu His Val Gln Glu Thr Lys Phe Gln Met Phe Gln Leu Ile Asp Ile Ala Arg Gln Thr Ala Gln Gly Met Asp Tyr Leu His Ala Lys Asn Ile Ile His Arg Asp Met Lys Ser Asn Asn Ile Phe Leu His Glu Gly Leu 480

Thr Val Lys Ile Gly Asp Phe Gly Leu Ala Thr Val Lys Ser Arg Trp 495

Ser Gly Ser Gln Gln Val Glu Gln Pro Thr Gly Ser Val Leu Trp Met 500 505 510

Ala Pro Glu Val Ile Arg Met Gln Asp Asn Asn Pro Phe Ser Phe Gln 515 520 525

Ser Asp Val Tyr Ser Tyr Gly Ile Val Leu Tyr Glu Leu Met Thr Gly 530 540

Glu Leu Pro Tyr Ser His Ile Asn Asn Arg Asp Gln Ile Ile Phe Met 545 550 560

Val Gly Arg Gly Tyr Ala Ser Pro Asp Leu Ser Lys Leu Tyr Lys Asn 565 570 575

Cys Pro Lys Ala Met Lys Arg Leu Val Ala Asp Cys Val Lys Lys Val 580 585

Lys Glu Glu Arg Pro Leu Phe Pro Gln Ile Leu Ser Ser Ile Glu Leu 595 600 605

Leu Gln His Ser Leu Pro Lys Ile Asn Arg Ser Ala Ser Glu Pro Ser 610 620

Leu His Arg Ala Ala His Thr Glu Asp Ile Asn Ala Cys Thr Leu Thr 625 630 635 640

Thr Ser Pro Arg Leu Pro Val Phe 645

WHAT IS CLAIMED IS:

 A method of identifying an individual at an increased risk for developing cancer, comprising:

amplifying a region of the c-raf-1 gene of said individual;

analyzing products of said amplification for evidence of mutation; and

classifying an individual having one or more mutations in said region as having an increased risk for developing cancer.

- 2. The method according to claim 1, wherein said region encodes at least amino acids 514 to 535 of SEQ ID NO:12.
- 3. The method according to claim 2, wherein said region encodes at least amino acids 500 to 550 of SEQ ID NO:12.
- 4. The method according to claim 3, wherein said region encodes at least amino acids 450 to 630 of SEQ ID NO:12.
- 5. The method according to claim 1, wherein said products are analyzed by DNA sequencing.
- 6. The method according to claim 1, wherein said amplification is effected using a polymerase chain reaction (PCR).
- 7. The method according to claim 6, wherein said PCR employs a primer comprising SEQ ID NO:7 and a primer comprising SEQ ID NO:8.
- 8. A method for determining a prognosis in patients afflicted with cancer, comprising:

amplifying a region of the c-raf-1 gene of said
individual;

analyzing products of said amplification for evidence of mutation; and

classifying a patient having no mutation in said region as being less likely to suffer disease relapse or having an increased chance of survival than a patient having one or more mutations in said region.

- 9. The method according to claim 8, wherein said region encodes at least amino acids 514 to 535 of SEQ ID NO:12.
- 10. The method according to claim 9, wherein said region encodes at least amino acids 500 to 550 of SEQ ID NO:12.
- 11. The method according to claim 10, wherein said region encodes at least amino acids 450 to 630 of SEQ ID NO:12.
- 12. The method according to claim 9, wherein said products are analyzed by DNA sequencing.
- 13. The method according to claim 9, wherein said amplification is effected using polymerase chain reaction (PCR).
- 14. The method according to claim 13, wherein said PCR employs a primer comprising SEQ ID NO:7 and a primer comprising SEQ ID NO:8.
- 15. A method for determining the proper course of treatment for a patient afflicted with cancer, comprising:

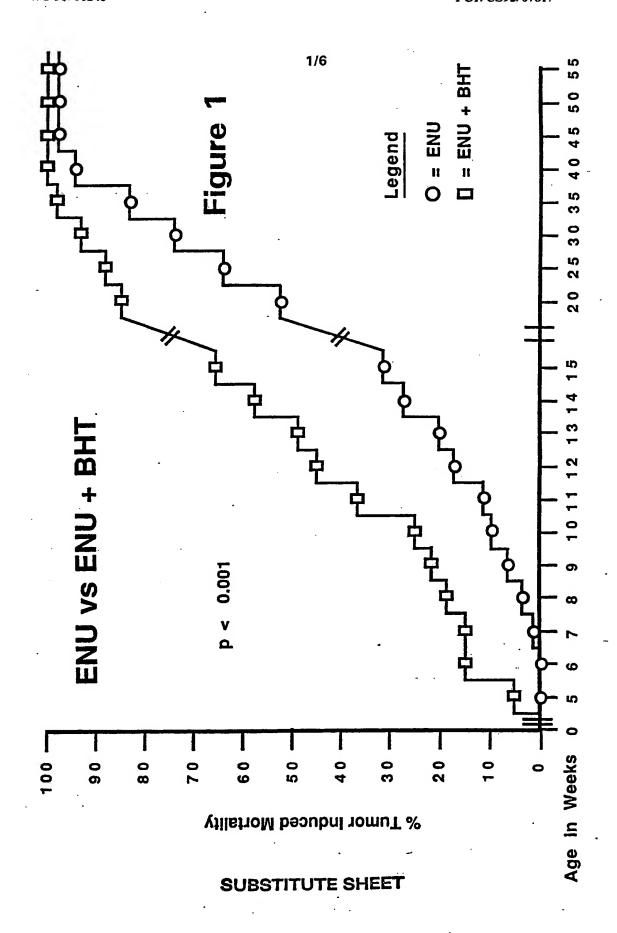
amplifying a region of the c-raf-1 gene of said
patient;

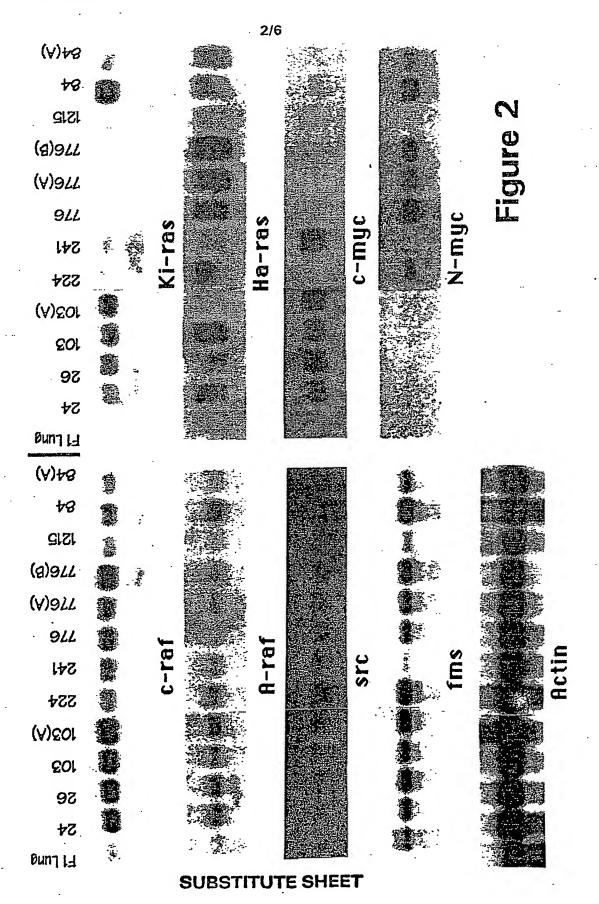
analyzing products of said amplification for evidence of mutation;

identifying a patient having at least one mutation in said region, which patient may require treatment proper for a patient having a lesser chance of survival or decreased time to relapse; and

identifying a patient lacking mutations in said region, which patient may require treatment proper for a patient having a greater chance of survival or being less likely to suffer disease relapse.

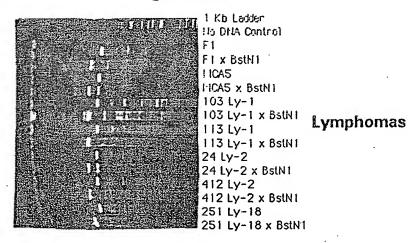
- 16. The method according to claim 15, wherein said region encodes at least amino acids 514 to 535 of SEQ ID NO:12.
- 17. The method according to claim 16, wherein said region encodes at least amino acids 500 to 550 of SEQ ID NO:12.
- 18. The method according to claim 17, wherein said region encodes at least amino acids 450 to 630 of SEQ ID NO:12.
- 19. The method according to claim 16, wherein said products are analyzed by DNA sequencing.
- 20. The method according to claim 16, wherein said amplification is effected using a polymerase chain reaction (PCR).
- 21. The method according to claim 20, wherein said PCR employs a primer comprising SEQ ID NO:7 and a primer comprising SEQ ID NO:8.

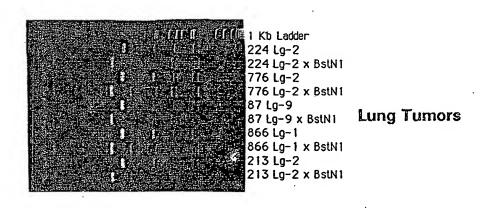


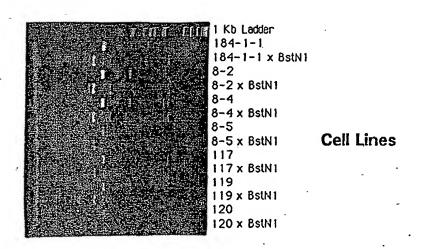


3/6

Figure 3







SUBSTITUTE SHEET

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Figure 4

pBR322 + Haelll

Probe

tRNA

v-raf

NFS/AKR F1

24 LY-1

26 LG-1

103 LG-1

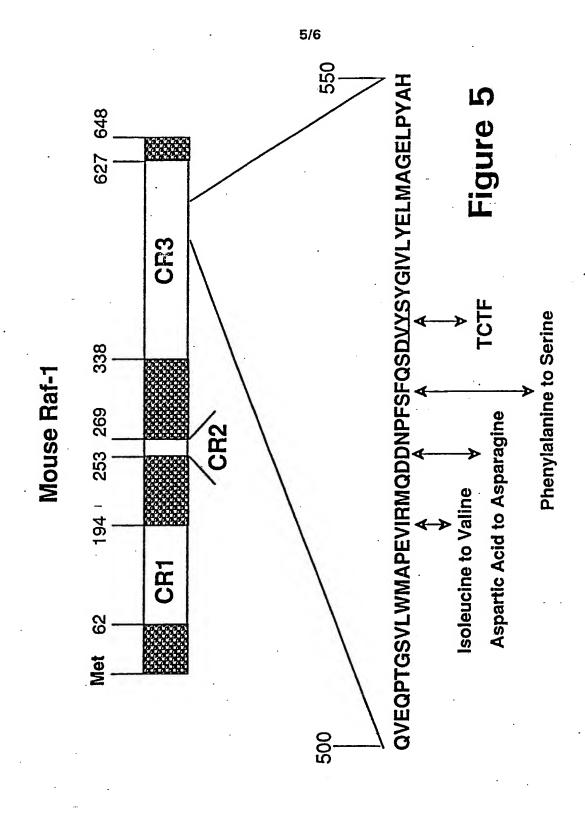
776 LG-2

145 LG-3

776 LG-3

776 LG-3A

SUBSTITUTE SHEET



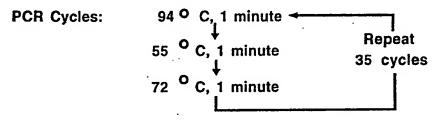
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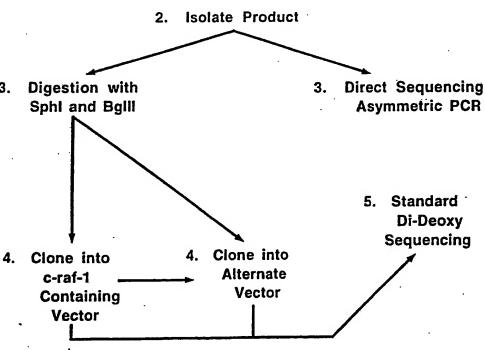
Figure 6

1. Polymerase Chain Reaction (PCR) Amplification of Target DNA

Either Genomic DNA or cDNA







SUBSTITUTE SHEET

INTERNATIONAL SEARCH REPORT

International Application No. PCT/US 92/07817

l. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) ⁶								
Accordin		tional Patent Classification (IPC) or to both h	tational Classification and IPC	İ				
11.03.	- 12 Q	1,00	·					
II. FIELD	S SEARCH	ED Minimum Docume	estation Sparrhed ⁷					
Classificati	oo System		Classification Symbols					
		7						
			•					
IPC5		C 12 Q						
-			r than Minimum Documentation is are included in Fields Searched [®]					
III. DOCU	MENTS CO	DISIDERED TO BE RELEVANT						
Category *	Çitəti	on of Document, ¹¹ with indication, where ap	propriate, of the relevant passages ¹²	Relevant to Claim No.13				
A		, 9112343 (CETUS CORPORAT August 1991,	ION)	1-21				
	se	e the whole document						
			·					
A	HE	, 9105064 (USA, REPR. BY ALTH/HUMAN S.) 18 April 1 e the whole document		1-21				
			•					
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A		, 9009456 (VIKTOR BALAZS) e the whole document	23 August 1990,	1-21				
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		•						
A	19	al Abstracts, volume 104, 86, (Columbus, Ohio, US), .: "The complete coding s	Bonner Tom I. et	1-21				
		f oncogene and the corres						
	th	e c-raf-1 gene", see page	141, abstract	j				
		2785c, & Nucleic Acids Re 09-1015	s. 1986, 14(2),					
"Special categories of cited documents: 10 T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the								
"E" earlier document but published on or after the international "X" document of particular relevance, the claimed invention filling date								
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another which is cited to establish the publication of another which is called to establish the publication of particular relevance, the claimed invention and the publication of particular relevance, the claimed invention and the publication of particular relevance, the claimed invention and the publication of particular relevance, the claimed invention and the publication of particular relevance.								
"O" document referring to an oral disclosure, use, exhibition or other means								
"P" document published prior to the international filing date but "&" document member of the same patent family later than the priority date claimed								
IV. CERTIFICATION Date of the Actual Completion of the International Search Date of Mailing of this International Search .								
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INTERNATIONAL SEARCH REPORT

International application No.

Pc:/US 92/07817

Box I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)	
This into	rnational search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:	
2 📗	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely. Remark: Claims 1-21 Diagnostic methods, c.f. PCT Rule 39(iv). Nevertheless, a search has been made concerning the subject matter of the application (the analytical method). Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such	
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(2).	
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)	
This Int	ernational Searching Authority found multiple inventions in this international application, as follows:	
1.	As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.	
2.	As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.	
3.	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:	
4.	No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:	
Remark	The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.	

ANNEX TO THE INTERNATIONAL SEARCH REPORT ON INTERNATIONAL PATENT APPLICATION NO.PCT/US 92/07817

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This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on 02/12/92. The European Patent office is in no way liable for these particulars which are merely given for the purpose of information.

cit	Patent document ed in search report	Publication date	Paten men	Publication date	
WO-A1-	9112343	22/08/91	AU-D- EP-A-	7758991 0514501	03/09/91 25/11/92
WO-A1-	9105064	18/04/91	AU-D- CA-A- EP-A-	6606190 2067114 0494968	28/04/91 03/04/91 22/07/92
WO-A1-	9009456	23/08/90	EP-A-	0458831	04/12/91
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